Yellow passion fruit tree sexual propagation by different mucilage extraction methods and substrates

Adriane Marinho de Assis^{1,*}, Sergio Ruffo Roberto², Vitor Camargo do Nascimento Júnior², Carla Liege Lonardoni Gomes de Oliveira², Rodrigo Thibes Hoshino², César Hideki Mashima², Celso Ramos² and Elder Andreazzi²

¹Universidade Federal de Pelotas, Departamento de Fitotecnia, CP 354, CEP 96010-900, Pelotas, RS, Brazil. ²Universidade Estadual de Londrina, Departamento de Agronomia, CP 10.011, CEP 86057-970, Londrina, PR, Brazil. *Corrresponding author, E-mail: agroadri17@gmail.com

ABSTRACT

86

The objective of this work was to evaluate the initial development of yellow passion fruit, as a function of the method of extracting the mucilage of the seeds (washing in water, fermentation in water and fermentation in water + sugar) and substrates (carbonized rice husk, coconut fiber and vermiculite). The completely randomized design with nine treatments and five replicates, with 50 seeds was used, in a 3 x 3 factorial arrangement (three mucilage extraction methods and three types of substrates). After 56 days of sowing, the following variables were evaluated seedling emergence percentage, number of leaves, leaf area, stem length and root largest length, dry mass of shoot and roots; and substrates characteristics such as pH, electric conductivity, density and water retention capacity. The emergence speed index was evaluated daily from sowing. It is verified that, except for the number of leaves and the shoot dry matter mass, the other variables were influenced by the study factors. In general, in the seeds without fermentation and fermentation in water mixed with sugar, the highest averages were obtained, regardless of the substrate used. Thus, both mucilage extraction methods and all tested substrates are indicated in the initial development of yellow passion fruit.

Key words: Passiflora edulis, initial development, carbonized rice husk, coconut fiber, vermiculite.

INTRODUCTION

The passion fruit plant (*Passiflora* sp.) from the Passifloraceae family has 630 species, 95% of them originated from South America (Lima and Cunha 2004). Brazil is considered the largest producer and consumer of passion fruit worldwide, with 703.489 tons collected in 2016 (Morgado et al., 2015; IBGE 2018).

One of the most important aspects for the success in the production areas is related to the use of quality seedlings. In the case of passion fruit, despite the cuttings (Sousa et al., 2014), grafting (Alexandre et al., 2004; Meletti 2011) and micro-propagation (Santos et al., 2010) being the options for getting seedlings, sexual propagation has been widely used (Wagner Júnior et al., 2007; Aguiar et al., 2014; Silva et al., 2015; Santos et al., 2016).

Knowledge about the seeds germination process of several *Passiflora* species is essential for propagation (Passos et al., 2004). Among the factors that can interfere with the seeds germination process are water, temperature and relative air humidity (Fachinello et al., 2005). However, passion fruit has a mucilage that involves the seeds creating a physical barrier and growth regulating substances, which can affect the process of germination (Pereira and Dias 2000) and promote the development of microorganisms.

Several authors have tested methods to extract mucilage from passion fruit seeds (Cardoso et al., 2001; Martins et al., 2006; Aguiar et al., 2014; Silva et al., 2015). However, there are no reports on the fermentation of these seeds in were mixed with sugar, technique used by Lone et al. (2014) with the same purpose in an experiment with pitaya (*Hilocereus* sp.), which reaffirms the need for studies to investigate its efficacy in passion fruit plant propagation.

Another factor to be considered is the substrate, medium in which roots of plants cultivated outside the soil develop (Kämpf 2005). Materials such vermiculite, coconut fiber and carbonized rice husk were tested in fruit species propagation (Pelizza et al., 2011; Yamamoto et al., 2013; Hussain et al., 2014). However, substrate selection or materials combination must be done according to the species at hand, in function of the specific physical and chemical characteristics of each material (Fachinello et al., 2005; Faria et al., 2010; Ristow et al., 2012). It is also necessary to check the availability and material cost since such requirements can be decisive in choosing the substrate.

Based on these aspects, the adoption of adequate procedures to separate seed from mucilage can represent a promising alternative for the propagation of passion fruit plant seeds and facilitate fruit handling during this operation. In addition, Wagner Júnior et al. (2006) emphasize that, this culture establishment success depends on several factors such as the use of an adequate substrate that may have great influence on the development of good quality seeds.

The objective of this work was to evaluate the influence of methods to extract mucilage from seeds and the use of different substrates in the sexual propagation of the yellow passion fruit tree.



MATERIALS AND METHODS

The experiment was conducted from September to November, 2012, at the Fruit Farming Sector of the Agronomy Department from Londrina State University Center for Agrarian Sciences, Paraná (latitude 23°23 S, longitude 51°11 O and altitude of 566 m).

Yellow passion fruit (*Passiflora edulis* Sims. f. *flavicarpa* Degener) seeds were used to study methods of mucilage extraction (washing with running water, water fermentation and fermentation in water + sugar) and substrates (carbonized rice husk, coconut fiber Amafibra[®] patter 47 and vermiculite medium granules). The experimental design was entirely randomized in a 3 x 3 factorial scheme (three mucilage extraction methods and three types pf substrates), with nine treatments and five replications of 50 seeds.

Fruit that showed complete maturation were collected from a single harvest lot. After the harvest, they were sectioned transversally and the seeds submitted to mucilage extraction by the fermentation method in water and water mixed with sugar (25 g L^{-1}). To do so, the experiment used a plastic recipient with capacity for 1.0 L, where seeds remained for four days. Next, they were washed with running water on a polyethylene sieve.

After the washing, seeds were placed immediately to dry away from the sun, at room temperature ($22 \text{ °C} \pm 0.5$), for four days, on a paper towel sheet. Then, the aryl was removed from seeds, rubbing them with a flannel towel. Next, they were stored in kraft paper bags, 0.48mm thick and kept in a refrigerator at 10 °C for 10 days (Aguiar et al., 2014). After this interval, seeding was realized on expandable polystyrene trays with 128 alveoli, with two seeds per alveolus, 1.0 cm deep in the referred substrates.

The trays remained in a nebulization chamber under a controlled intermittent regime by a timer and solenoid valve, programmed to nebulize for 10 seconds at each three-minute interval. The nebulizer nozzle used (Mist Dan Sprinklers model, Israel) has a flow of 35 liters per hour. The nebulization chamber is inserted in the agricultural greenhouse covered with a transparent polyethylene film and sombrite fabric 30%. Then, when seedlings were 50 cm high, they were thinned and the most vigorous plant remained.

During the experiment, the average temperature registered in the interior of the greenhouse was $23 \text{ }^{\circ}\text{C} \pm 2.0$ and the average air relative humidity was $57\% \pm 0.8$. Germination was evaluated after seeding by emerged seedlings percentage. Daily counting after seeding was carried out from emergence to stabilization, to assess emergence speed index. Emergence speed index was determined according to Maguire (1962).

The following variables were evaluated 56 days after seeding (Figure 1): emergence percentage, number of leaves, leaf area (cm²), stem and greater root length (cm) and aerial part and roots dry matter mass per plant (g). Plants with open cotyledons were considered emerged (Lima et al., 2006). Leaf area determination was done from images obtained by a scanner and later by the the SisCob images analysis program, developed by Embrapa CNPDIA.



Figure 1. Yellow passion fruit tree (*Passiflora edulis*) seedlings 56 days after experiment installation. Londrina, PR, Brazil.

Stem and greater root length were measures by a graded ruler, using the distance between the neck and the apex of the seedling stem and root, respectively, as reference. During the determination of the roots and aerial part dry matter mass, they were placed to dry in a greenhouse with forced air circulation at 60 °C, until reaching constant weight. In addition, emergence speed index was evaluated daily from seeding.

Electrical conductivity (μ S cm⁻¹), density (kg m⁻³), pH and substrates water retention capacity (mL kg⁻¹) were determined according to Kämpf et al. (2006).

Data were submitted to analysis of variance and the means compared by the Tukey test at 5% of probability. For variables originated from the percentages, data were transformed according to the equation for $\frac{1}{\sqrt{x+100}}$; and for the variables originated from the counting, data were transformed according to equation $\frac{1}{\sqrt{x+1}}$.

RESULTS AND DISCUSSION

With the exception of number of leaves, stem length and aerial part dry matter mass, the other variables were affected by the study factors (Tables 1 and 2).

As for emergence percentage, in regards to mucilage extraction methods, there was no significant difference, regardless the substrate. However, in regards to substrates, the coconut fiber showed higher seeds water fermentation mean, differing statistically from the other materials (Table 1).

A research work on the propagation of yellow passion fruit trees, Cardoso et al. (2001) found that seeds fermentation promoted greater emergence percentage. Similarly, Lopes et al. (2007) observed greater germination percentage (84.5%) in seeds submitted to total aryl removal.

Table 1. Emergence percentage (EP %), emergency speed index (ESI), number of leaves per plant (NL) and
leaf area per yellow passion plant (LA (cm ²)), in function of mucilage extraction methods (MEM) and
substrates. Londrina, PR, Brazil. 2012.

	MEM	Substrates						
		CRI	CRH		F	V	VE	
	NF	78,40	Aa	77,20	Aa	68,40	Aa	
EP (%)	FW	66,40	Ba	80,00	Aa	64,80	Ba	
	FWS	77,20	Aa	84,00	Aa	76,40	Aa	
	CV (%)=6,10	Value of <i>I</i>	P(M)=0,024	V	alue of	<i>P</i> (S)=0,007		
	NF	1,75	Aab	1,72	Aa	1,45	Ab	
ESI	FW	1,45	Bb	1,91	Aa	1,45	Вb	
	FWS	1,92	Aa	2,11	Aa	1,80	Aa	
	CV (%)=15,37	Value of F	P(M)=0,002	2 Value of P (S)=0,004				
	NF	2,79	Aa	2,79	Aa	3,14	Aa	
NL	FW	3,20	Aa	2,91	Aa	3,33	Aa	
	FWS	3,20	Aa	3,03	Aa	3,10	Aa	
	CV (%)=3,75 Value of P (M)=0,079		Value of P (S)=0,060					
	NF	3,17	Aa	3,42	Aa	4,10	Aa	
LA (cm ²)	FW	3,92	ABa	3,26	Ba	4,30	Aa	
	FWS	3,72	Aa	3,83	Aa	3,82	Aa	
	CV (%)=16,73	Value of F	P(M)=0,462	V	alue of	<i>P</i> (S)=0,039		

Means followed by the same lowercase letters in the column do not differ statistically among them by the Tukey test (P<0.05). Legend: CRH: carbonized rice husk; CF: coconut fiber; VE: vermiculite; NF: no fermentation (water washing); FW: fermentation in water; FWS: fermentation in water + sugar; (MEM): mucilage extraction methods.

According to Wagner Júnior et al. (2006), seeds germination is influenced by substrates properties such water retention capacity, aeration, among others. In the present study, one factor that could have influenced emergence was the coconut fiber greater water retention capacity (Table 3), considering that the water plays a fundamental role in triggering germination process metabolic activities (Fachinello et al., 2005) and that, in some cases, an additional supply of substrate water for seeds germination (Martins et al., 2009).

	MEM			Subst	rates		10	
		CRH		C	CF		3	
	NF	3.21	Aa	3.31	Aa	3.24	Aa	
SL (cm)	FW	3.42	Aa	3.60	Aa	3.53	Aa	
	FWS	3.39	Aa	3.58	Aa	3.52	Aa	
CV	(%)=8,85	Value of .	P (M)=	0.407	Value	of P (S)=0.374		
	NF	0.595	Aa	0.740	Aa	0.715	Aa	
APMM (g)	FW	0.574	Aa	0.761	Aa	0.733	Aa	
	FWS	0.740	Aa	0.893	Aa	0.787	Aa	
CV ((%)=24,92	Value of J	P (M)=	0.122	Value	of P (S)=0.057		
	NF	5.33	Aab	5.63	Aa	5.45	Aa	
GRL (cm)	FW	5.18	Ba	5.60	Aa	5.13	Aa	
	FWS	5.97	Aa	5.96	Aa	5.40	Aa	
CV	Value of $P(M)=0.019$ Value of $P(S)=0.051$							
	NF	0.225	Aab	0.291	Aa	0.245	Aa	
RMM (g)	FW	0.189	Вb	0.347	Aa	0.267	ABa	
	FWS	0.339	Aa	0.427	Aa	0.302	Aa	
CV((%)=30,63	Value of 1	P (M)=	0.007	Value	of P (S)=0.007		

Means followed by the same uppercase letters in the line and lowercase in the column show no statistical difference among them by the Tukey test (P < 0.05). Legend: CRH: carbonized rice husk; CF: coconut fiber; VE: vermiculite; NF: no fermentation (water washing); FW: fermentation in water; FWS: fermentation in water + sugar; (MEM): mucilage extraction methods.

Table 3. pH, electrical conductivity (EC μ S cm⁻¹), density (kg m⁻³) and water retention capacity (WRC mL kg⁻¹) of carbonized rice husk substrates (CRH), coconut fiber (CF) and vermiculite (VE), 56 days after seeding. Londrina, PR, Brazil. 2012.

Substrates	pН	EC	Density	WRC
	-	μS cm ⁻¹	kg m ⁻³	mL kg ⁻¹
CRH	7.43	223.40	157.06	371.00
CF	6.70	266.60	109.90	760.00
VE	7.77	257.40	282.04	170.00

As for the emergence speed index, in regards to mucilage extraction methods, the highest mean was obtained from fermentation with water and sugar, showing no difference from the seeds without fermentation by using carbonized rice husk; however, there were no significant differences between the methods in coconut fiber while fermentation in water and sugar showed the highest vermiculite mean. In regards to substrates, coconut fiber showed the highest mean compared to the other materials, only for water fermentation (Table 1).

Cardoso et al. (2001) verified that fermentation promoted yellow passion fruit tree seedlings emergence sooner when compared to those originated from seeds without fermentation; while Aguiar et al. (2014), in an experiment with the same fruit tree observed that the emergence speed was superior when seeding was realized in coconut fiber and carbonized rice husk.

Martins et al. (2009) evaluated the sexual propagation of the pupunha palm tree (*Bactris gasipaes*) mentioned that the fast and uniform germination of the seeds, followed by prompt emergence are desirable during seedlings development, since the longer the seedling remains in the initial development stadia the more vulnerable it will become to the environment's adverse conditions. In addition to the factors studied in the present experiment, it is possible to infer that the temperature also influenced the time required for germination and the IVE, since it was in agreement with the values recommended for *Passiflora edulis* seeding (Ministério da Agricultura, Pecuária e Abastecimento 2009).

In regards to number of leaves, there were no statistical differences related to mucilage removal methods and substrates. However, the largest leaf area was register for vermiculite for seeds whose fermentation done in water, differing from the other substrates (Table 1). According to Cavalcante et al. (2009), during the photosynthesis period, the largest leaf area is more efficient in transporting mineral and organic solutes to vegetal tissues.

In regards to stem length and aerial part dry matter, there was no significant difference between mucilage extraction methods and substrates (Table 2). In a study carried out by Silva et al. (2010), the highest means for passion fruit tree seedlings height were observed in recipients with soil + manure, which did not differ significantly from the mix of soil + 150 mg dm⁻³ of simple superphosphate and Plantmax[®].

Great root length per plant showed significant differences in regards to mucilage extraction methods only for carbonized rice husk, with fermentation with water mixed with sugar showing the greatest length the greater length, not differing statistically from the method without fermentation (Table 2).

In regards to roots dry matter mass, the extraction methods analysis showed that plants originated from seeds with water with sugar fermentation presented higher means, showing no difference from those submitted to fermentation in carbonized rice husk. On the other hand, substrates in coconut fiber showed higher mean only when submitted to water fermentation, with no difference from vermiculite (Table 2).

Findings from this study may be the result of the coconut fiber low density when compared to vermiculite and carbonized rice husk (Table 3), considering the high-density values may represent greater resistance to roots expansion in the substrate. According to Carrijo et al. (2002), the good physical properties of the coconut fiber highlight the possible use of this substrate.

In regards to the substrates chemical properties, the pH varied from 6.70 to 7.77 and such values did not interfere in the initial development of the seedlings (Table 3). Electrical conductivity, although the highest mean was found in coconut fiber (Table 3), all values are classified as an ideal for substrates from seedbeds and tray seedlings, since, according to Ballester-Olmos (1993), values between 200-350 μ S cm⁻¹ are appropriate for most plants. Negreiros et al. (2004) reported that a good substrate offers adequate conditions for germination and root system development of seedlings.

For the production of passion fruit tree seedlings Costa et al. (2009) recommend the mix of vermiculite with soil and an organic compound. However, in the present study, in general, both treatment without fermentation and fermentation in water with sugar showed the highest means regardless of the substrate used. However, greater practicality was observed during the removal of the mucilage after fermentation in water + sugar. In regards to substrates, the grower can opt for the most cost-effective material available in the region, considering that this is one of the most relevant items in seedlings production. In addition, agricultural residues destination represents an environmental gain in function of their reduction in the nature.

CONCLUSIONS

Mucilage extraction without fermentation and with fermentation in water mixed with sugar, as well as coconut fiber substrates, carbonized rice husk and vermiculite are recommended for yellow passion fruit tree sexual propagation.

ACKNOWLEDGEMENTS

To CAPES (Coordination for the Improvement of Higher Education Personnel) and CNPq (National Council for Scientific and Technological Development) for the financial support.

REFERENCES

Aguiar RS, Yamamoto LY, Preti EA, Souza GRB, Sbrussi CAG, Oliveira EAP, Assis AM, Roberto SR and Neves CSVJ (2014) Extração de mucilagem e substratos no desenvolvimento de plântulas de maracujazeiro-amarelo. Semina: Ciências Agrárias 35: 605-612.

Alexandre RS, Wagner Júnior A, Negreiros JRS, Parizzoto A and Bruckner CH (2004) Germinação de sementes de genótipos de maracujazeiro. Pesquisa Agropecuária Brasileira 39: 1239-1245.

Ballester-Olmos JF (1993) Substratos para el cultivo de plantas ornamentales. Madrid, Saijen, 44p.

Cardoso GD, Tavares JC, Ferreira RLF, Câmara FAA and Carmos GA (2001) Desenvolvimento de mudas de maracujazeiro-amarelo obtidas de sementes extraídas por fermentação. Revista Brasileira de Fruticultura 23: 639-642.

Carrijo OA, Liz RS and Makishima N (2002) Fibra da casca do coco verde como substrato agrícola. Horticultura Brasileira 20: 533-535.

Cavalcante LF, Silva GF, Gheyi HR, Dias TJ, Alves JC and Costa APM (2009) Crescimento de mudas de maracujazeiro amarelo em solo salino com esterco bovino líquido fermentado. Revista Brasileira de Ciências Agrárias 4: 414-420.



Costa E, Rodrigues ET, Alves VB, Dos Santos LCR and Vieira LCR (2009) Efeitos da ambiência, recipientes e substratos no desenvolvimento de mudas de maracujazeiro-amarelo em Aquidauana - MS. Revista Brasileira de Fruticultura 31: 236-244.

Fachinello JC, Hoffmann A and Nachtigal JC (2005) Propagação de plantas frutíferas. Embrapa Informações Tecnológicas, Brasília, 221p.

Faria RT, Assis AMA and Fiúza JRPC (2010) Cultivo de Orquídeas. Mecenas, Londrina, 208p.

Hussain I, Assis, AM, Yamamoto LY, Koyama R and Roberto SR (2014) Indole butyric acid and substrates influence on multiplication of blackberry 'Xavante'. Ciência Rural 44: 1761-1765.

IBGE (2018) Sistema IBGE de Recuperação automática. Tabela 1613-Área destinada à colheita, área colhida, quantidade produzida, rendimento médio e valor da produção das lavouras permanentes. https://sidra.ibge.gov.br/tabela/1613#resultado. Acesso 2 jan 2018.

Kämpf AN (2005) Produção comercial de plantas ornamentais. Agrolivros, Guaíba, 256p.

Kämpf AN, Takane RJ and Siqueira PTV (2006) Floricultura: Técnicas de preparo de substratos. LK Editora e Comunicação, Brasília, 132p.

Lima AA and Cunha MAP (2004) Maracujá: Produção e qualidade na Passicultura. Embrapa Mandioca e Fruticultura, Cruz das Almas, 396p.

Lima AA, Caldas RC and Santos VS (2006) Germinação e crescimento de espécies de maracujá. Revista Brasileira de Fruticultura 28: 125-127.

Lone AB, Colombo RC, Favetta V, Takahashi LSA and Faria RT (2014) Temperatura na germinação de sementes de genótipos de pitaya. Semina: Ciências Agrárias 35: 2251-2258.

Lopes JC, Bono GM and Alexandre VMM (2007) Germinação e vigor de plantas de maracujazeiro 'amarelo' em diferentes estádios de maturação do fruto, arilo e substrato. Ciência e Agrotecnologia 31: 1340-1346.

Maguire JD (1962) Speed of germination-aid in selection and evaluation for seedling emergence and vigor. Crop Science 2: 176-177.

Martins CC, Bovi MLA and Spiering SH (2009) Umedecimento do substrato na emergência e vigor de plântulas de pupunheira. Revista Brasileira de Fruticultura 31: 224-230.

Martins MR, Reis MC, Neto JAM, Gusmão LL and Gomes JJA (2006) Influência de diferentes métodos de remoção do arilo na germinação de sementes de maracujazeiro-amarelo (*Passiflora edulis* Sims F. *flavicarpa* Deg.). Revista da FZVA 13: 28-38.

Meletti LMM (2011) Avanços na cultura do maracujá no Brasil. Revista Brasileira de Fruticultura vol.especial: 83-91.

Ministério da Agricultura, Pecuária e Abastecimento (2009) Regras para análise de sementes. Secretaria de Defesa Agropecuária, Brasília, 399p.

Morgado MAD, Bruckner CH, Rosado LDS and Santos CEM (2015) Desenvolvimento de mudas de maracujazeiro-azedo enxertadas em espécies silvestres de *Passiflora*. Revista Brasileira de Fruticultura 37: 471-479.

Negreiros JRS, Álvares VS, Braga LR and Bruckner CH (2004) Diferentes substratos na formação de mudas de maracujazeiro-amarelo.Revista Ceres 51: 243-249.

Passos IRS, Matos GVC, Meletti LMM, Scott MDS, Bernacci LC and Vieira MAR (2004) Utilização do ácido giberélico para a quebra de dormência de sementes de *Passiflora nitida* Kunth germinadas *in vitro*. Revista Brasileira de Fruticultura 26: 380-381.

Pelizza TR, Damiani CR, Rufato AR, Souza ALK, Ribeiro MF and Schuch MW (2011) Microestaquia em mirtileiro com diferentes porções do ramo e substratos. Bragantia 70: 319-324.

Pereira KJC and Dias DCFS (2000) Germinação e vigor de sementes de maracujá-amarelo (*Passiflora edulis* Sims. f. *flavicarpa* Deg.) submetidas a diferentes métodos de remoção da mucilagem. Revista Brasileira de Sementes 22: 288-291.

Ristow NC, Antunes LEC and Carpenedo S (2012) Substratos para o enraizamento de microestacas de mirtileiro cultivar Georgiagem. Revista Brasileira de Fruticultura 34: 262-268.

Santos CHB, Neto AJC, Junghans TG, Jesus ON and Girardi EA (2016) Estádio de maturação de frutos e influência de ácido giberélico na emergência e crescimento de *Passiflora* spp. Revista Ciência Agronômica 47: 481-490.

Santos FC, Ramos JD, Pasqual M, Rezende JC, Santos FC and Villa F (2010) Micropropagação do maracujazeirodo-sono. Revista Ceres 57: 112-117.

Silva EA, Maruyama WI, Mendonça V, Francisco MGS, Bardiviesso DM and Tosta MS (2010) Composição de substratos e tamanho de recipientes na produção e qualidade das mudas de maracujazeiro 'amarelo'. Ciência e Agrotecnologia 34: 588-595.

Silva SM, Oliveira RC, Almeida RF, Júnior AS and Santos CM (2015) Aryl removal methods and passion fruit seed positions: Germination and emergence. Journal of Seed Science 37: 125-130.

Sousa CM, Santos MP and Carvalho BM (2014) Enraizamento de estacas de maracujazeiro-doce (*Passiflora alata* Curtis). Científica 42: 68-73.

Wagner Júnior A, Alexandre RS, Negreiros JRS, Pimentel LD, Costa Silva JO and Bruckner CH (2006) Influência do substrato na germinação e desenvolvimento inicial de plantas de maracujazeiro amarelo (*Passiflora edulis* Sims f. *flavicarpa* Deg.). Ciência e Agrotecnologia 30: 643-647.

Wagner Júnior A, Dos Santos CE, Alexandre RS, Costa and Silva JO, Negreiros JRS, Pimentel LD, Álvares VS and Bruckner CH (2007) Efeito da pré-embebição das sementes e do substrato na germinação e no desenvolvimento inicial do maracujazeiro-doce. Revista Ceres 54: 001-006.

Yamamoto LY, Koyama R, Borges WFS, Antunes LEC, Assis AM and Roberto SR (2013) Substratos no enraizamento de estacas herbáceas de amora-preta Xavante. Ciência Rural 43: 15-20.

Received: December 22, 2017. Accepted: February 21, 2018. Published: May 11, 2018.